

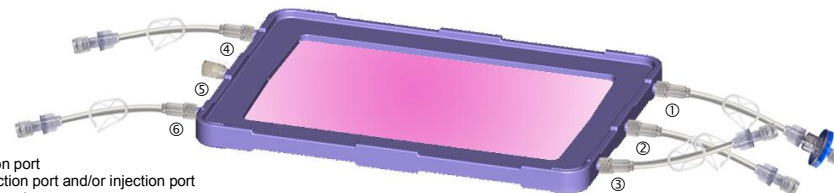
Recovery of adherent cells

1. Check all the clamps are closed.
2. Hold the **CLINicell**[®] vertically, the ventilation port (1) directed to the top.
3. Remove the ventilation port (1) and the extraction port (6) caps.
4. Connect a 50 ml syringe to the extraction port (6).
5. Open the extraction port (6) and the ventilation port (1) clamps.
6. Ensure the ventilation port (1) is raised enough to allow air to get into the **CLINicell**[®] and aspirate the supernatant.
7. Close the extraction port (6) and ventilation port (1) clamps.
8. Disconnect the syringe (create a forced vacuum) and recover the supernatant.
9. Repeat steps 4 to 8 as necessary.
10. Wash the **CLINicell**[®] once or twice (refer to the [washing procedure](#) as explained in the previous section).
11. Remove the ventilation port (1) and injection port (3) caps. (if necessary)
12. Connect to the injection port (3) a 20 ml syringe containing the dissociation solution (e.g. 10 ml of trypsin /EDTA) and 10 ml of air.
13. Open the ventilation port (1) and injection (3) port clamps.
14. Lift the syringe vertically and inject the dissociation solution.
15. Close the ventilation port (1) clamp and secure the cap.
16. Close the injection port (3) clamp, disconnect the syringe (create a forced vacuum) and secure the cap.
17. Gently agitate the **CLINicell**[®] to homogenise the repartition of the solution.
18. Incubate at 37°C if necessary.
19. Hold the **CLINicell**[®] vertically, the ventilation port (1) directed to the top.
20. Remove the ventilation port (1) and injection port (3) caps.
21. Connect to the injection port (3) a 50 ml syringe containing the medium (or an other solution) used to stop the dissociation reaction and 10 ml of air.
22. Open the injection port (3) and ventilation port (1) clamps.
23. Lift the syringe vertically and inject the medium.
24. Close the ventilation port (1) clamp and gently agitate the **CLINicell**[®] to homogenise.
25. Open the ventilation port (1) clamp.
26. Aspirate the whole suspension ensuring the ventilation port (1) is raised enough to allow air to get into the **CLINicell**[®].
27. Close the ventilation port (1) clamp and replace the cap.
28. Close the injection port (3) clamp, disconnect the syringe (create a forced vacuum), recover the cell suspension and secure the cap.
29. If necessary, wash the **CLINicell**[®] once or twice (Refer to the [washing procedure](#) as explained in the previous section).
30. Throw the used **CLINicell**[®] in an appropriate bin.

CLINicell[®] 250

Medical Device

Sterile single use cell culture cassette



- ① ventilation port
- ② air extraction port and/or injection port
- ③ injection port
- ④,⑤ extraction ports
- ⑥ injection site latex free

User Instructions.

Warning : please read this user manual prior any manipulation

I. Storage precautions :

Store in a clean, dry and dark location, at room temperature.

II. Handling precautions :

- a - **Do not use CLINicell[®] if it is damaged. Do not reuse.**
Always verify packaging integrity. We guarantee **CLINicell**[®] sterility if packaging is unopened and intact.
- b - **Manipulate CLINicell[®] in a sterile environment.**
Avoid contact between the access ports and your hands or the flow hood working surface. Check all the ways are closed or connected before getting **CLINicell**[®] out of the flow hood.
- c - Check the clamps are closed before removing the caps.
- d - For any manipulation (injection or extraction of fluids) always open the ventilation port (1) to avoid overpressure within **CLINicell**[®]. Be careful not to wet the filter.
- e - We suggest using "Luer Lock" syringe, as this allows the syringe to securely screw to the access port.
- f - For safety, we recommend using blunt-end needles to aspirate solutions with the syringe.
- g - To limit the contamination risks when you remove the syringe from an injection/extraction port, always create a forced vacuum to draw any residual liquid remaining in the port end.

Manipulation of the **CLINicell[®] 250 Culture Cassette**

Recommendations :

- We recommend to prepare the cell suspension within the optimal volume of medium (125 ml) and to inject the whole suspension into the **CLINicell[®]**.
- For high concentration cultures (from 1.10^6 cells/ml) of non-adherent cells, we recommend to re-suspend the cells each two days, gently agitate the **CLINicell[®]** or gently tap on the rigid frame.

Filling of the **CLINicell[®] 250 Culture Cassette**

1. Close all the clamps.
2. Prepare the cell suspension in the optimal volume of medium (125 ml)
3. Hold the **CLINicell[®]** vertically, the injection port (3) directed to the top.
4. Remove the caps from the ventilation (1) and injection (3) ports.
5. Take the cell suspension using a 50 ml syringe containing about 10 ml of air. (Once the suspension has been injected, the air will ensure in-flowing cell suspension remaining in the injection way.)
6. Connect the syringe to the injection port (3).
7. Open the injection (3) and ventilation (1) port clamps.
8. Lift the syringe to inject the cell suspension down into the injection port (3).
9. Close the injection (3) and ventilation (1) port clamps and remove the syringe (create a forced vacuum).
10. Repeat steps 5 to 9 to fill the cassette with 125 ml of cell suspension (whilst injecting the cell suspension ensure the ventilation port (1) is raised to allow air to escape the system).
11. Once the cell suspension has been injected, close the ventilation port (1) clamp and secure the cap.
12. Close the injection port (3) clamp, remove the syringe (create a forced vacuum) and secure the cap.
13. Remove air extraction (2) port cap, connect the syringe and open the clamp.
14. Raise the **CLINicell[®]** vertically, aspirate the residual air from cassette letting the way empty, and close the clamp.
15. Remove the syringe (create a forced vacuum) and secure the air extraction port (2) cap.
16. Incubate the **CLINicell[®]**.

Recovery of non-adherent cells

1. Check all the clamps are closed.
2. Re-suspend the cells, gently agitate the **CLINicell[®]** or gently tap on the rigid frame.
3. Hold the **CLINicell[®]** vertically, the ventilation port (1) directed to the top.
4. Remove the ventilation port (1) and the extraction port (6) caps.
5. Connect a 50 ml syringe to the extraction port (6).
6. Open the ventilation port (1) and extraction port (6) clamps.
7. Aspirate the cell suspension (ensure the ventilation port (1) is raised enough to allow air to get into the system).
8. Close the ventilation port (1) and extraction port (6) clamps.
9. Disconnect the syringe (create a forced vacuum) and recover the cell suspension.
10. Repeat steps 5 to 9 to recover the total cell suspension.
11. Close the ventilation port (1) clamp and secure the cap.
12. Close the extraction port (6) clamp, disconnect the syringe (create a forced vacuum) and replace the cap.

Recommended washing procedure following recovery of culture from your **CLINicell[®] 250**

1. Hold the **CLINicell[®]** vertically, the injection port (3) directed to the top.
2. Check all the clamps are closed and remove the caps from the ventilation port (1) and injection port (3).
3. Connect a 50 ml syringe, containing the washing solution (e.g. 20 ml of PBS) and 10 ml of air, to the injection port (3).
4. Open the ventilation port (1) and the injection port (3) clamps.
5. Lift the syringe vertically and inject the washing solution.
6. Close the ventilation port (1) clamp and gently agitate the **CLINicell[®]**.
7. Hold the **CLINicell[®]**, ventilation port (1) directed to the top, and open the clamp.
8. Ensure the ventilation port (1) is raised enough to allow air to get into the **CLINicell[®]** and aspirate the washing solution.
9. Close the ventilation port (1) clamp and secure the cap.
10. Close the injection port (3) clamp, disconnect the syringe (create a forced vacuum), recover the washing solution and replace the cap.